

天青石蓝苏木素染色试剂盒

货号: G4470

规格: 2×50mL/2×100mL

保存: 2-8°C, 避光保存, 有效期 1 年。

产品组成:

| 名称 | 2×50mL | 2×100mL | 保存 |
|---------------------|--------|---------|-----------|
| 试剂(A): 天青石蓝 B 染色液 | 50mL | 100mL | 2-8°C, 避光 |
| 试剂(B): Mayer 苏木素染色液 | 50mL | 100mL | 2-8°C, 避光 |

产品介绍:

单纯的明矾苏木素细胞核染色后的缺点是对随后应用的任何酸性染料比较敏感, 最常见于 Van Gieson 染色和 masson 三色染色。Van Gieson 染色时使用苦味酸品红可脱掉大部分苏木素的颜色, 以致细胞核几乎看不到, 在这种情况下使用铁苏木素或天青石蓝可抵抗苦味酸的影响而达到满意效果。目前, 可靠的方法是天青石蓝和明矾苏木素联合使用, 其优势在于天青石蓝能抗酸, 天青石蓝溶液中的高铁盐可加强明矾苏木素与细胞核的结合力, 使细胞核染色更深, 因此可以达到抗酸的目的。

操作步骤: (仅供参考)

1. 石蜡切片脱蜡, 梯度乙醇水化至水。
2. 切片滴加天青石蓝染色液染色 2-3min, 蒸馏水冲洗 5-10s。
3. 切片滴加 Mayer 苏木素染色液染色 2-3min, 切片自来水洗 10min 蓝化。(见注意事项 2)
4. 按实验要求继续进行。

注意事项:

1. 随着染色液放置时间的延长, 染色力会有所下降, 染色时间也需延长。染液的使用程度越强, 染色力下降越快, 所需染色时间也就越长。冰冻切片的染色时间应缩短, 在无缓冲液的福尔马林溶液浸泡时间较长的组织及脱钙组织染色时时间应延长。
2. 亦可使用蓝化液处理 2-3min 代替, 蓝化液可使用 0.2~1% 氨水 (G1822) 或 Scott 促蓝液或 0.1~1% 碳酸锂溶液 (G1841)。
3. 为了您的安全和健康, 请穿实验服并戴一次性手套操作。





Celestine Blue Hematoxylin Stain Kit

Cat: G4470

Size: 2×50mL/2×100mL

Storage: 2-8°C, avoid light, valid for 1 year.

Kit Components

| Reagent | 2×50mL | 2×100mL | Storage |
|--|--------|---------|--------------------|
| Reagent(A): Celestine blue B solution | 50mL | 100mL | 2-8°C, avoid light |
| Reagent(B): Mayer Hematoxylin Solution | 50mL | 100mL | 2-8°C, avoid light |

Introduction

The disadvantage of pure alum hematoxylin dyeing is that it is more sensitive to any acid dyes used later. It is most common in Van Gieson staining and Masson Trichrome staining. Using fuchsin picrate when Van Gieson's staining removes most of the hematoxylin color so that the nucleus is barely visible. In this case, the use of cyrtoxylin or lapis lazuli blue can resist the influence of picric acid and achieve satisfactory results. At present, the reliable method is the combination of lapis lazuli blue and aluminum hematoxylin. Its advantage is that the ferrate in lapis lazuli blue solution can strengthen the binding force between alumosin and nucleus, and make the nucleus stain deeper to achieve the purpose of acid resistance.

Protocol(for reference only)

1. Dewax the paraffin section and hydration by series ethanol.
2. Add Celestine blue B solution for 2-3min. Rinse with distilled water for 5-10s.
3. Add Mayer Hematoxylin Solution for 2-3min. Bluing with tap water for 10-20min. (Refer Note 2)
4. Operate the next steps according to the experimental process.

Note

1. With the prolongation of the time of putting the dye solution, the dyeing power will be decreased and the dyeing time will be prolonged. The stronger the dye is used, the faster the dyeing power decreases and the longer the dyeing time is required. Slice dewaxing should be as clean as possible. Series ethanol should be replaced frequently.
2. The bluing Solution can be replaced by 0.2-1% ammonia water (G1822) or Scott blue promoting liquid or 0.1-1% lithium carbonate solution (G1841).
3. To prevent over staining, the dyeing time of frozen section must be short. The staining time should be prolonged for the tissues and decalcified tissues soaked in formalin without buffer.
4. For your safety and health, please wear experimental clothes and disposable gloves.

